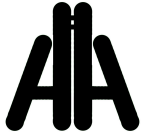

*AccuEZ*TM Chloramphenicol Fast Detection ELISA Kit

User Guide

(Catalog No. AAF-21121)

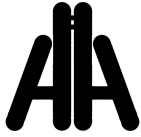
Enzyme-Linked Immunosorbent Assay Kit for Detection of
Chloramphenicol

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General Description

Chloramphenicol is a broad-spectrum antibiotic effective against a wide variety of Gram-positive and Gram-negative bacteria. It is no longer commonly prescribed to patients because of the presence of resistant bacteria and safety concerns. One of its most prominent adverse effects is bone marrow toxicity, which includes direct bone marrow suppression and aplastic anemia, with the latter being relatively rare but unpredictable, not dosage-dependent, and generally fatal. Chloramphenicol is especially toxic for neonates. However, due to its low cost and effectiveness to improve yield, Chloramphenicol has been illegally used in aquaculture and therefore is present in some seafood such as shrimp or crawfish. There are reports on its presence in milk, honey, and meat products as well.

The AccuAffinity *AccuEZ*TM Chloramphenicol ELISA Kit can **quickly**, **sensitively**, and **accurately** determine the presence of Chloramphenicol in animal tissue (e.g. chicken, beef, fish, or shrimp), honey, or milk, providing a vital tool to prevent consumption of food tainted with Chloramphenicol.

Intended Use

The AccuAffinity *AccuEZ*TM Chloramphenicol ELISA Kit utilizes competitive ELISA for the quantitative and qualitative analysis of Chloramphenicol in food samples including animal tissue (e.g. chicken, beef, fish, or shrimp), honey and milk. The limit of detection (LOD) of Chloramphenicol is 0.05 ppb (0.05 ng/mL)

Safety Instructions

To receive complete safety information on this product, contact AccuAffinity, Inc. and request Material Safety Data Sheets. Stop solution is 1N sulfuric acid. Handle with care.

Assay Principle

*AccuEZ*TM Chloramphenicol kit is a competitive enzyme-labeled immunoassay. Each well of the 96-well plate has been pre-coated with anti-Chloramphenicol antibody. During assay, Chloramphenicol standard solution or samples are added to test wells, followed by



adding horseradish peroxidase (HRP)-Chloramphenicol conjugate, which will compete with Chloramphenicol in standard/sample for binding to antibody during 30-minute incubation. After washing the plate, a clear HRP substrate is added to the wells leading to a colored product, and optical density is inversely related to Chloramphenicol concentration in the samples. The accurate concentration of Chloramphenicol can then be determined by interpolation using the standard curve constructed in the same run.

Reagents and Materials Provided

The reagents included in the kit are sufficient for performing 96 measurements (including standards and samples). Reagents and materials in each kit include:

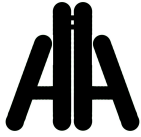
- a) 1 microtiter plate containing 12 test strips of 8 wells sealed in an aluminized pouch with desiccant.
- b) 6 vials each containing 0.5 mL of Chloramphenicol standard with 0, 0.05, 0.15, 0.45, 1.35, 4.05 ng/mL of Chloramphenicol respectively.
- c) 1 vial containing 0.1 mL Chloramphenicol-HRP conjugate (100×).
- d) 1 bottle containing 10ml sample diluent (1 x)
- e) 1 bottle containing 10ml assay buffer (1 x)
- f) 1 bottle containing 15 mL microtiter plate wash solution (20×).
- g) 1 bottle containing 11 mL substrate (1×).
- h) 1 bottle containing 11 mL stop solution (1×).
- i) 2 microtiter plate sealers.
- j) 1 booklet of instruction.

Materials Required but not Provided

- a) Microplate reader with 450 nm filter.
- b) Pipet capable of dispensing 20-200 μ l

Assay Procedure

- Equilibrate kit components at room temperature (20-25°C) for at least 30 min prior to running the test, and thoroughly mix all liquid components before use.
- Use test strips as needed on the frame, and store unused strips in the re-closable bag at 2-8°C.
- Number standards and samples according to positions on microtiter plate. All standards and samples need duplicate measurement for accuracy.



Sample preparation:

Meat (including fish and shrimp) samples need to be processed as followings before ELISA assay:

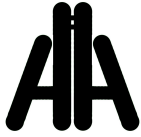
1. Homogenize sample
2. Weight 1.0 g homogenized sample
3. Add 2ml Ethyl Acetate, mix by vortex thoroughly
4. Centrifuge 4000rpm for 10 min at room temperature
5. Take 1 ml supernatant
6. Evaporate to dryness in a nitrogen evaporator
7. Add 0.5 ml hexane to reconstitute
8. Add 0.5 ml of sample diluent
9. Vortex vigorously
10. Centrifuge 4000rpm for 10 min at room temperature
11. Discard upper layer, take 50 μ l bottom aqueous phase to ELISA plate

ELISA assay:

1. Prepare Wash Solution by diluting 1 part of Wash Solution Concentrate (20x) with 19 parts of distilled water to Wash Solution (1x)
2. Prepare HRP working solution by diluting 1 part of HRP conjugate (100x) with 99 parts of Assay Buffer to HRP working solution (1x)
3. Add Chloramphenicol standard sample, or unknown samples, 50 μ l/well in duplicate
4. Add HRP working solution 50 μ l/well, gently shake plate by hand for 1 min, and incubate at room temperature for 30 min
5. Wash plate 4 times with wash solution (1x), 200 μ l/well wash buffer each time
6. Add 100 μ l/well TMB-Ultra solution, and incubate plate at room temperature for 15 min in dark place
7. Add 100 μ l/well stopping solution, read OD_{450nm}

Calculating Results

Two approaches can be used to obtain Chloramphenicol concentration results from the assay. Semi-quantitative results can be obtained with the first approach while quantitative results can be calculated with the second. Please note the negative correlation between absorbance reading (OD₄₅₀) and Chloramphenicol concentration in the sample.



1. Semi-quantitative detection of Chloramphenicol

A simple comparison of average sample absorbance to absorption of standards will give the range of Chloramphenicol concentration (ng/mL or ppb) in the samples. For example, Sample 1 has an average absorbance of 0.6, and Sample 2 of 1.2, and OD₄₅₀ of standard solutions are as follows: 2.300 for 0 ng/mL; 2.000 for 0.05 ng/mL; 1.650 for 0.15 ng/mL; 0.950 for 0.45 ng/mL; 0.400 for 1.35 ng/mL; 0.165 for 4.05 ng/mL. It is immediately known that Chloramphenicol concentration of Sample 1 is between 0.45 ng/mL and 1.35 ng/mL, while Sample 2 contains 0.15-0.45 ng/mL Chloramphenicol.

2. Quantitative calculation of Chloramphenicol concentration

a) Calculate B/B₀

Dividing average absorbance of each standard and sample (B) by absorbance of first standard (the standard with 0 ng/mL Chloramphenicol concentration, B₀) to obtain percentage absorbance.

$$\text{percentage absorbance (\%)} = B/B_0 \times 100\%$$

B — average absorbance of a standard or sample

B₀ — average absorbance of 0 ng/mL standard

b) A standard curve is obtained by graphing the percentage absorbance of standards (Y axis) versus their corresponding concentration (X axis) on semi-log graph paper (which should be a linear relationship), and sample concentration can be read from this standard curve. Alternatively, Chloramphenicol concentration in the samples can be calculated with regression equation correlating percentage absorbance to Chloramphenicol concentration. Graphing software can also be used for quick analyses of large number of samples.

Performance Data

Range of Standard Curve:	0 – 4.05 ng/ml
Assay Quantitative Range:	0.05 – 4.05 ng/ml
Assay Time:	40 min

Limit of Detection (LOD):

Meat (after calculation of dilution factor) 0.1 ppb

Recovery: 70-130%



Sensitivity (defined as the average of absorbance from 6 zero-standards minus 3 times of standard deviation): 0.05 ng/ml

Precision:

Intra-assay CV <10%

Inter-assay CV <15%

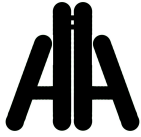
Precautions

1. Assay kit should be stored at 2-8°C and avoid freezing conditions; unused test strips should be sealed in reclosable bag; colorless substrate is sensitive to light so prolonged exposure to light needs to be avoided.
2. Reagents should be brought to room temperature (20-25°C) prior to use. A room temperature of lower than 20°C or failure to equilibrate reagents or samples to room temperature could result in low OD readings for all samples. All reagents should be put back into 2-8°C storage immediately after use.
3. Adhere to assay protocol on reaction temperature and time, and use pipet to add components whenever possible. Results are solely based on OD450 readings from plate/strip reader.
4. Reagents need to be thoroughly mixed to improve reproducibility.
5. During all incubation steps, avoid light and seal plate with sealer.
6. If wells are dried out during plate wash steps, linearity of standard curve will be negatively affected and reproducibility will be poor. Therefore, substrate addition should be carried out immediately after tapping the plate dry (following the last wash).
7. The stop solution is 1N sulfuric acid. Avoid contact with skin or clothing. Immediately clean up any spills and wash area with copious amounts of water. If contact should occur, immediately flush with copious amounts of water.
8. Do not use reagents beyond expiration date. Dilution or adulteration of reagents or samples not called for in the procedure may result in adverse changes in sensitivity and OD reading. Do not substitute reagents from kits with different lot numbers.
9. Obvious color in substrate suggests expiration and it should be discarded. When absorbance of zero-standard is lower than 0.8, the reagents may have expired.

Storage and Expiration Date

Storage: All components of the kit should be stored at 2-8°C.

Expiration Date: This kit expires 12 months after manufacturing date.



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Technical Assistance

For ordering or technical assistance regarding this kit, or for additional information about AccuAffinity products, please email support@accuaffinity.com or call (408) 368-1364.

General Limited Warranty

AccuAffinity, Inc. warrants its manufactured products against defects in materials and workmanship when used in accordance with the applicable instructions for a period not to extend beyond a product's printed expiration date. AccuAffinity makes no other warranty, expressed or implied. There is no warranty of merchantability or fitness for a particular purpose. The warranty provided herein and the data, specifications and descriptions of AccuAffinity products appearing in published catalogues and product literature may not be altered except by express written agreement signed by an officer of AccuAffinity. Representations, oral or written, which are inconsistent with this warranty or such publications are not authorized and, if given, should not be relied upon.

In the event of a breach of the foregoing warranty, AccuAffinity Inc.'s sole obligation shall be to repair or replace, at its option, any product or part thereof that proves defective in materials or workmanship within the warranty period, provided the customer notifies AccuAffinity promptly of any such defect. The exclusive remedy provided herein shall not be deemed to have failed of its essential purpose so long as AccuAffinity is willing and able to repair or replace any nonconforming AccuAffinity product or part. AccuAffinity shall not be liable for consequential, incidental, special or any other indirect damages resulting from economic loss or property damage sustained by a customer from the use of its products. However, in some states the purchaser may have rights under state law in addition to those provided by this warranty.

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